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(54) **SUGAR TRANSPORT SEQUENCES, YEAST STRAINS HAVING IMPROVED SUGAR UPTAKE, AND METHODS OF USE**

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This patent is subject to a terminal disclaimer.

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Related U.S. Application Data

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(60) Provisional application No. 61/061,417, filed on Jun. 13, 2008.

(51) **Int. Cl.**
C12P 7/06 (2006.01)
C07K 14/39 (2006.01)
C12P 7/18 (2006.01)

(52) **U.S. Cl.**
CPC . *C07K 14/39* (2013.01); *C12P 7/06* (2013.01); *Y02E 50/17* (2013.01); *C12P 7/18* (2013.01)
USPC **435/161**

(58) **Field of Classification Search**
CPC *C07K 14/39*; *C12P 7/18*
USPC **435/161**
See application file for complete search history.

(56) **References Cited**

U.S. PATENT DOCUMENTS

8,105,811 B2 * 1/2012 Jeffries et al. 435/161

* cited by examiner

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(57) **ABSTRACT**

Disclosed are nucleic acid constructs comprising coding sequences operably linked to a promoter not natively associated with the coding sequence. The coding sequences encode *Pichia stipitis* proteins that allow recombinant strains of *Saccharomyces cerevisiae* expressing the protein to grow on xylose, and allow or increase uptake of xylose by *Pichia stipitis* or *Saccharomyces cerevisiae* expressing the coding sequences. Expression of the coding sequences enhances uptake of xylose and/or glucose, allowing increased ethanol or xylitol production.

12 Claims, 8 Drawing Sheets

ClustalW (v1.8) multiple sequence alignment

3 Sequences Aligned Alignment Score = 34327
Gaps Inserted = 0 Conserved Identities = 100%

Pairwise Alignment Mode: Slow
Pairwise Alignment Parameters:
Open Gap Penalty = 10.0 Extend Gap Penalty = 0.1

Multiple Alignment Parameters:
Open Gap Penalty = 10.0 Extend Gap Penalty = 0.1
Delay Divergent = 40% Transitions: Weighted

Processing time: 3.3 seconds

SUT4 ORF 1 ATGTCCCTCACAAGATTTACCCCTCGGGTCTCAAACCCCAATCGATGGTTC 50
(SSEQ ID NO:3)
SUT2.nt.mv 1 ATGTCCCTCACAAGATTTACCCCTCGGGTCTCAAACCCCAATCGATGGTTC 50
(SSEQ ID NO:12)
sut3 1 ATGTCCCTCACAAGATTTACCCCTCGGGTCTCAAACCCCAATCGATGGTTC 50

SUT4 ORF 51 TTCCATCCCTCGAAGATAAAGTTGAGCAAAGTTGGTCCCTCAAATAGCCAAA 100
SUT2.nt.mv 51 TTCCATCCCTCGAAGATAAAGTTGAGCAAAGTTGGTCCCTCAAATAGCCAAA 100
sut3 51 TTCCATCCCTCGAAGATAAAGTTGAGCAAAGTTGGTCCCTCAAATAGCCAAA 100

SUT4 ORF 101 GTGATTTAGCTTCTATTTCCAGCAACAGGTATCAAAGCCCTATCTCTTGGTT 150
SUT2.nt.mv 101 GTGATTTAGCTTCTATTTCCAGCAACAGGTATCAAAGCCCTATCTCTTGGTT 150
sut3 101 GTGATTTAGCTTCTATTTCCAGCAACAGGTATCAAAGCCCTATCTCTTGGTT 150

SUT4 ORF 151 TGTTCCTTCTGCAATGTTGGTTGGCTTGGTGGCTTGGTATTGGSTTTTCCA 200
SUT2.nt.mv 151 TGTTCCTTCTGCAATGTTGGTTGGCTTGGTGGCTTGGTATTGGSTTTTCCA 200
sut3 151 TGTTCCTTCTGCAATGTTGGTTGGCTTGGTGGCTTGGTATTGGSTTTTCCA 200

SUT4 ORF 201 TACCGGTACTATTTCCGGTTTCTTAAATATGTTTGATTTCTTTCCAGAT 250
SUT2.nt.mv 201 TACCGGTACTATTTCCGGTTTCTTAAATATGTTTGATTTCTTTCCAGAT 250
sut3 201 TACCGGTACTATTTCCGGTTTCTTAAATATGTTTGATTTCTTTCCAGAT 250

SUT4 ORF 251 TTGGTCAAGATGGTTCTGGAAGGAAATATTTGTTCTGATATCAGAGTGGT 300
SUT2.nt.mv 251 TTGGTCAAGATGGTTCTGGAAGGAAATATTTGTTCTGATATCAGAGTGGT 300
sut3 251 TTGGTCAAGATGGTTCTGGAAGGAAATATTTGTTCTGATATCAGAGTGGT 300

SUT4 ORF 301 TTGATTTGTTTCCATTTTAAACATTTGGTTGTCGCAATGGTGGTATTTTCT 350
SUT2.nt.mv 301 TTGATTTGTTTCCATTTTAAACATTTGGTTGTCGCAATGGTGGTATTTTCT 350
sut3 301 TTGATTTGTTTCCATTTTAAACATTTGGTTGTCGCAATGGTGGTATTTTCT 350

SUT4 ORF 351 TTCTAAGATAGGAGATGTTTACGGTAGAAGAATGGTATCATTTTCAAGCTA 400
SUT2.nt.mv 351 TTCTAAGATAGGAGATGTTTACGGTAGAAGAATGGTATCATTTTCAAGCTA 400
sut3 351 TTCTAAGATAGGAGATGTTTACGGTAGAAGAATGGTATCATTTTCAAGCTA 400

SUT4 ORF 401 TGGTTGTCATAGTGGTGGTATTAATCATCCAGATCTCGTCCCAAGATAAG 450
SUT2.nt.mv 401 TGGTTGTCATAGTGGTGGTATTAATCATCCAGATCTCGTCCCAAGATAAG 450
sut3 401 TGGTTGTCATAGTGGTGGTATTAATCATCCAGATCTCGTCCCAAGATAAG 450

SUT4 ORF 451 TGGTACCAACTTACAATTTGGACGTTGAGGTTACAGGATTAGCTGTTGGTAC 500
SUT2.nt.mv 451 TGGTACCAACTTACAATTTGGACGTTGAGGTTACAGGATTAGCTGTTGGTAC 500

FIG. 1 (1 of 3)

seq3 451 TGGTACCAACTTACAAATYKACCTEGAGTTACAGGATTAAGCTGTTGGTAC 500

SEQ4 ORF 501 TGTTCGCTTCTGCTCCAAATSTTCAATTATGAAATGCTCCCAAGCATT 550
SEQ2 .nt .sw 501 TGTTCGCTGTTGCTCCCAATSTTCAATTATGAAATGCTCCCAAGCATT 550
seq3 501 TGTTCGCTGTTGCTCCCAATSTTCAATTATGAAATGCTCCCAAGCATT 550

SEQ4 ORF 551 TGAGAGGTACTTDEGTATACCTTTACCAATATATGCAATCACCTTAGGTATT 600
SEQ2 .nt .sw 551 TGAGAGGTACTTDEGTATACCTTTACCAATATATGCAATCACCTTAGGTATT 600
seq3 551 TGAGAGGTACTTDEGTATACCTTTACCAATATATGCAATCACCTTAGGTATT 600

SEQ4 ORF 601 TTCATTCGCTTACTTGTCTCACTTATGGAACCAAGATTTAAATGATTCAG 650
SEQ2 .nt .sw 601 TTCATTCGCTTACTTGTCTCACTTATGGAACCAAGATTTAAATGATTCAG 650
seq3 601 TTCATTCGCTTACTTGTCTCACTTATGGAACCAAGATTTAAATGATTCAG 650

SEQ4 ORF 651 ACAATGGAGAGTTCCCTTGGGTTATATTTTCCCTTGGCTATTTTCTTAG 700
SEQ2 .nt .sw 651 ACAATGAGAGATTCCTTGGGTTATATTTTCCCTTGGCTATTTTCTTAG 700
seq3 651 ACAATGAGAGATTCCTTGGGTTATATTTTCCCTTGGCTATTTTCTTAG 700

SEQ4 ORF 701 TTCTTGGTATGTTGGCTATATCCAGAATCCCAAGATTCCTTCAATGAAAG 750
SEQ2 .nt .sw 701 TTCTTGGTATGTTGGCTATATCCAGAATCCCAAGATTCCTTCAATGAAAG 750
seq3 701 TTCTTGGTATGTTGGCTATATCCAGAATCCCAAGATTCCTTCAATGAAAG 750

SEQ4 ORF 751 AAGAAATCCAGAGAGCCAGAGATCCCTTCCAGATCCCAAGATTTTTC 800
SEQ2 .nt .sw 751 AAGAAATCCAGAGAGCCAGAGATCCCTTCCAGATCCCAAGATTTTTC 800
seq3 751 AAGAAATCCAGAGAGCCAGAGATCCCTTCCAGATCCCAAGATTTTTC 800

SEQ4 ORF 801 TCCAGAGATCCAGGTTCTTACACTGAAATTCATTTGATTCAGCTGTA 850
SEQ2 .nt .sw 801 TCCAGAGATCCAGGTTCTTACACTGAAATTCATTTGATTCAGCTGTA 850
seq3 801 TCCAGAGATCCAGGTTCTTACACTGAAATTCATTTGATTCAGCTGTA 850

SEQ4 ORF 851 TTACAGAGAACTGCTCCAGGTTCTGCTTCTGGATGGAATGATTAAC 900
SEQ2 .nt .sw 851 TTACAGAGAACTGCTCCAGGTTCTGCTTCTGGATGGAATGATTAAC 900
seq3 851 TTACAGAGAACTGCTCCAGGTTCTGCTTCTGGATGGAATGATTAAC 900

SEQ4 ORF 901 GGTAAACCCGCTATTTTACAGAGATTTATCATGGAATTAATTTACAGTC 950
SEQ2 .nt .sw 901 GGTAAACCCGCTATTTTACAGAGATTTATCATGGAATTAATTTACAGTC 950
seq3 901 GGTAAACCCGCTATTTTACAGAGATTTATCATGGAATTAATTTACAGTC 950

SEQ4 ORF 951 TTTCACCAATTAACCTGCTGCACTATTTCTTCTATTAAGGAACTACAA 1000
SEQ2 .nt .sw 951 TTTCACCAATTAACCTGCTGCACTATTTCTTCTATTAAGGAACTACAA 1000
seq3 951 TTTCACCAATTAACCTGCTGCACTATTTCTTCTATTAAGGAACTACAA 1000

SEQ4 ORF 1001 TCTTCCAGGCTGTTGGTTTCCAGAAATTCCTTCCAGACTTCCATCATCTTA 1050
SEQ2 .nt .sw 1001 TCTTCCAGGCTGTTGGTTTCCAGAAATTCCTTCCAGACTTCCATCATCTTA 1050
seq3 1001 TCTTCCAGGCTGTTGGTTTCCAGAAATTCCTTCCAGACTTCCATCATCTTA 1050

SEQ4 ORF 1051 GGTACAGTCAACTTTCTTTCTACATTTGTTGGTATTTGGGCAATTHAAG 1100
SEQ2 .nt .sw 1051 GGTACAGTCAACTTTCTTTCTACATTTGTTGGTATTTGGGCAATTHAAG 1100
seq3 1051 GGTACAGTCAACTTTCTTTCTACATTTGTTGGTATTTGGGCAATTHAAG 1100

SEQ4 ORF 1101 ATTTGAGAGAGACAATGTTTGTAGTCCGTTCTGCTGGTATGTTGGT 1150
SEQ2 .nt .sw 1101 ATTTGAGAGAGACAATGTTTGTAGTCCGTTCTGCTGGTATGTTGGT 1150
seq3 1101 ATTTGAGAGAGACAATGTTTGTAGTCCGTTCTGCTGGTATGTTGGT 1150

FIG. 1 (2 of 3)

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SUT4 ORF      1151  GTTTCAGCATTACTTACTCCTGTCATTGGTACAACACTCTTTTGTTCATTGATGGA 1200
SUT2 .nt..mv  1151  GTTTCAGCATTACTTACTCCTGTCATTGGTACA&CTCATTTGTTCATTGATGGA 1200
sut.3         1151  GTTTCAGCATTACTTACTCCTGTCATTGGTACAACCTCTTTTGTTCATTGATGGA 1200
***** *
SUT4 ORF      1201  GTAGTACGATAACGACACACACCCCGTCA&CTGTCTGGAATGCTATGATCCT 1250
SUT2 .nt..mv  1201  GTAGTACGATAACGACACACACCCCGTCA&CTGTCTGGAATGCTATGATCCT 1250
sut.3         1201  GTAGTACGATAACGACACACACCCCGTCA&CTGTCTGGAATGCTATGATCCT 1250
*****
SUT4 ORF      1251  TATCACTTGTGTTTGTTCATCTTCTCTCTTTCCTGCTGACTTGGGCTGGAGGCTG 1300
SUT2 .nt..mv  1251  TATCACTTGTGTTTGTTCATCTTCTCTCTTTCCTGCTGACTTGGGCTGGAGGCTG 1300
sut.3         1251  TATCACTTGTGTTTGTTCATCTTCTCTCTTTCCTGCTGACTTGGGCTGGAGGCTG 1300
*****
SUT4 ORF      1301  TTTTTCAGATCATTTTCGAAATCATAATCCTTTCAGATCAGATCCAGGCT 1350
SUT2 .nt..mv  1301  TTTTTCAGATCATTTTCGAAATCATAATCCTTTCAGATCAGATCCAGGCT 1350
sut.3         1301  TTTTTCAGATCATTTTCGAAATCATAATCCTTTCAGATCAGATCCAGGCT 1350
*****
SUT4 ORF      1351  ATGTCATTTGCCACTGCGCGTAAACGATGATGAGGCTTCTTGTGATTTCAAT 1400
SUT2 .nt..mv  1351  ATGTCATTTGCCACTGCGCGTAAACGATGATGAGGCTTCTTGTGATTTCAAT 1400
sut.3         1351  ATGTCATTTGCCACTGCGCGTAAACGATGATGAGGCTTCTTGTGATTTCAAT 1400
*****
SUT4 ORF      1401  CTGCACTCCATTCATTTGTTAAAGCCACCA&CTTCA&ATTCGGCTTTGTGT 1450
SUT2 .nt..mv  1401  CTGCACTCCATTCATTTGTTAAAGCCACCA&CTTCA&ATTCGGCTTTGTGT 1450
sut.3         1401  CTGCACTCCATTCATTTGTTAAAGCCACCA&CTTCA&ATTCGGCTTTGTGT 1450
*****
SUT4 ORF      1451  TTACTGCTTGTGTTTACTCTTTTCGTTCTTCTATGCTACTCTTFTTTCAGC 1500
SUT2 .nt..mv  1451  TTACTGCTTGTGTTTACTCTTTTCGTTCTTCTATGCTACTCTTFTTTCAGC 1500
sut.3         1451  TTACTGCTTGTGTTTACTCTTTTCGTTCTTCTATGCTACTCTTFTTTCAGC 1500
*****
SUT4 ORF      1501  GAAACCAAAGGTTTFTCTTTCGAAAGAGTTGATGAGTTGTACCCCTGAGGG 1550
SUT2 .nt..mv  1501  GAAACCAAAGGTTTFTCTTTCGAAAGAGTTGATGAGTTGTACCCCTGAGGG 1550
sut.3         1501  G. A&CCAAAGGTTTFTCTTTCGAAAGAGTTGATGAGTTGTACCCCTGAGGG 1550
*****
SUT4 ORF      1551  AATTGACCCATGGAATTCGCTGCAATGCTTCTCTTCTTSCACACACAC 1600
SUT2 .nt..mv  1551  AATTGACCCATGGAATTCGCTGCAATGCTTCTCTTCTTSCACACACAC 1600
sut.3         1551  AATTGACCCATGGAATTCGCTGCAATGCTTCTCTTCTTSCACACACAC 1600
*****
SUT4 ORF      1601  AAATGCAAAACTCTACTTATGCTGCGCAAGCAAGAGACCAAGACCAAGTT 1650
SUT2 .nt..mv  1601  AAATGCAAAACTCTACTTATGCTGCGCAAGCAAGAGACCAAGACCAAGTT 1650
sut.3         1601  AAATGCAAAACTCTACTTATGCTGCGCAAGCAAGAGACCAAGACCAAGTT 1650
*****
SUT4 ORF      1651  TAG      1653  ++++  34 NT changes
SUT2 .nt..mv  1651  TAG      1653  ++    3 NT changes
sut.3         1651  TAGTT  1656  +++   6 NT changes
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FIG. 1 (3 of 3)

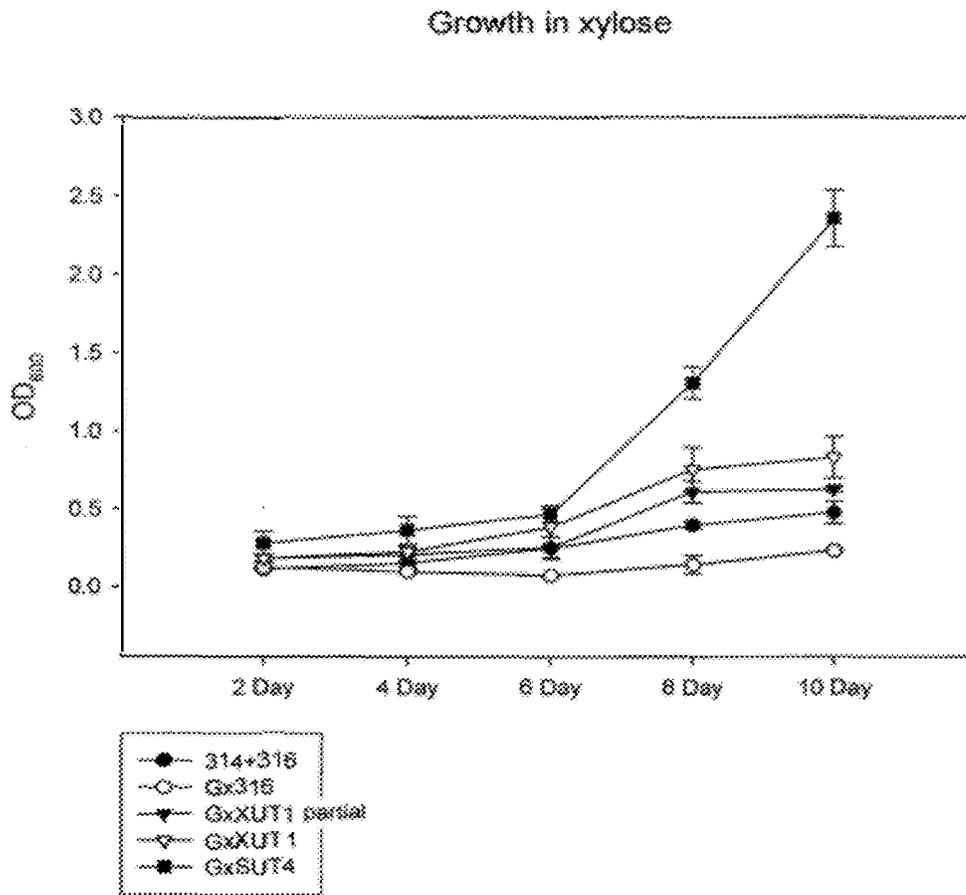


FIG. 2

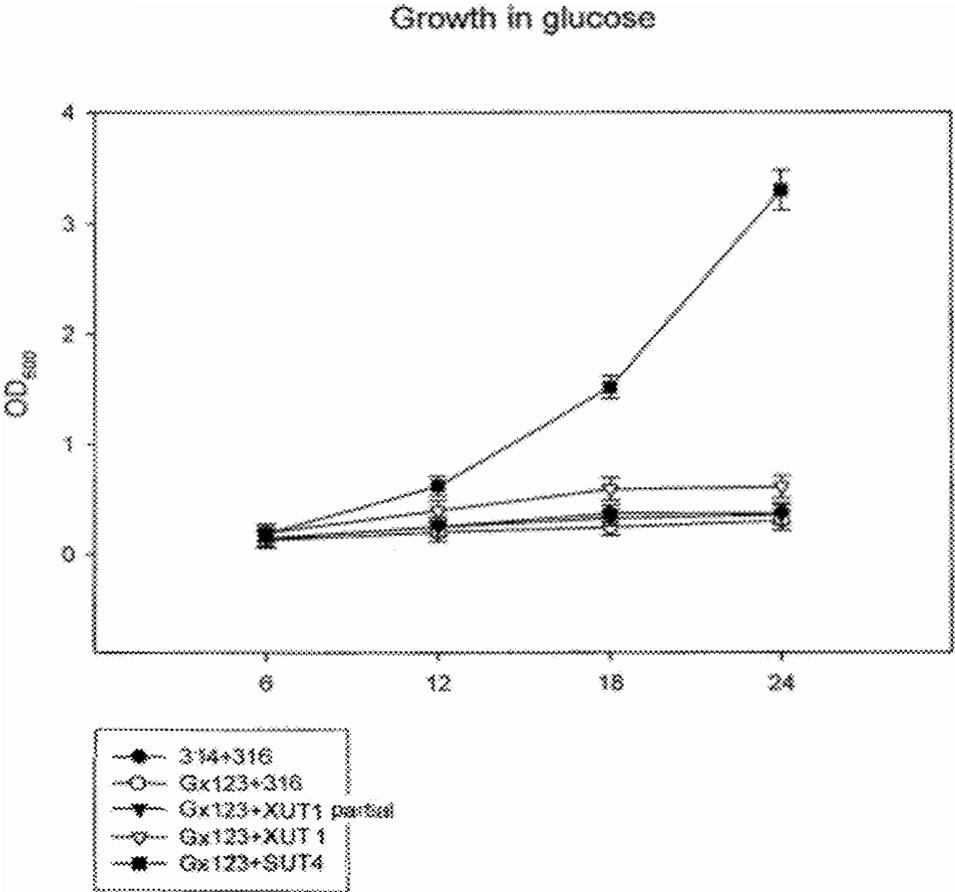


FIG. 3

50 mM ¹⁴C-glucose or xylose uptake with time course.

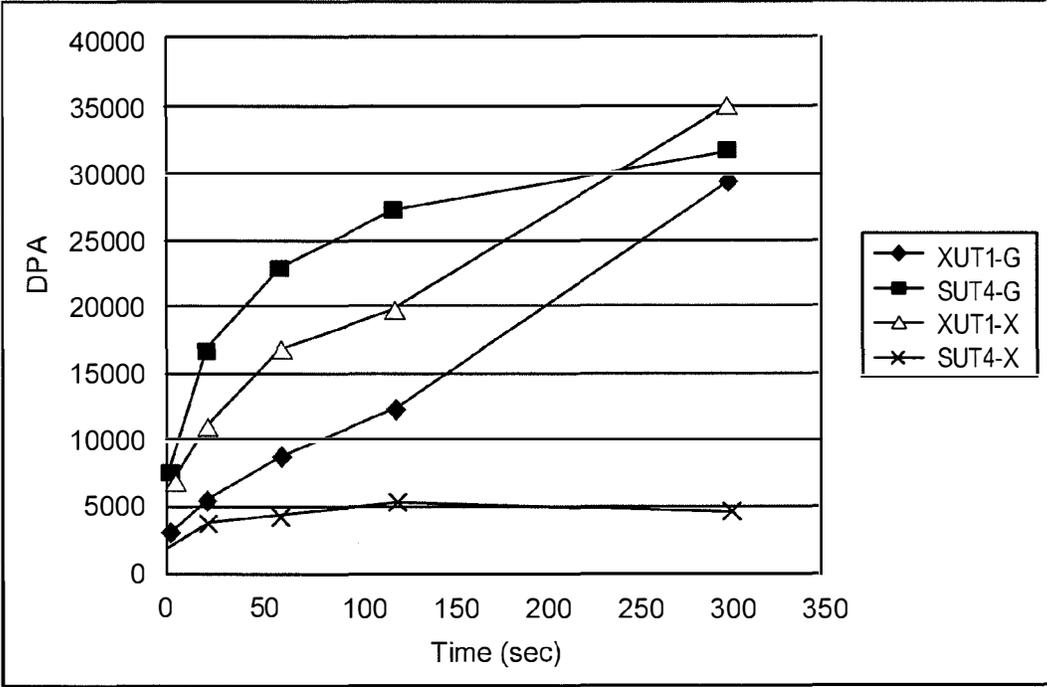


FIG. 4

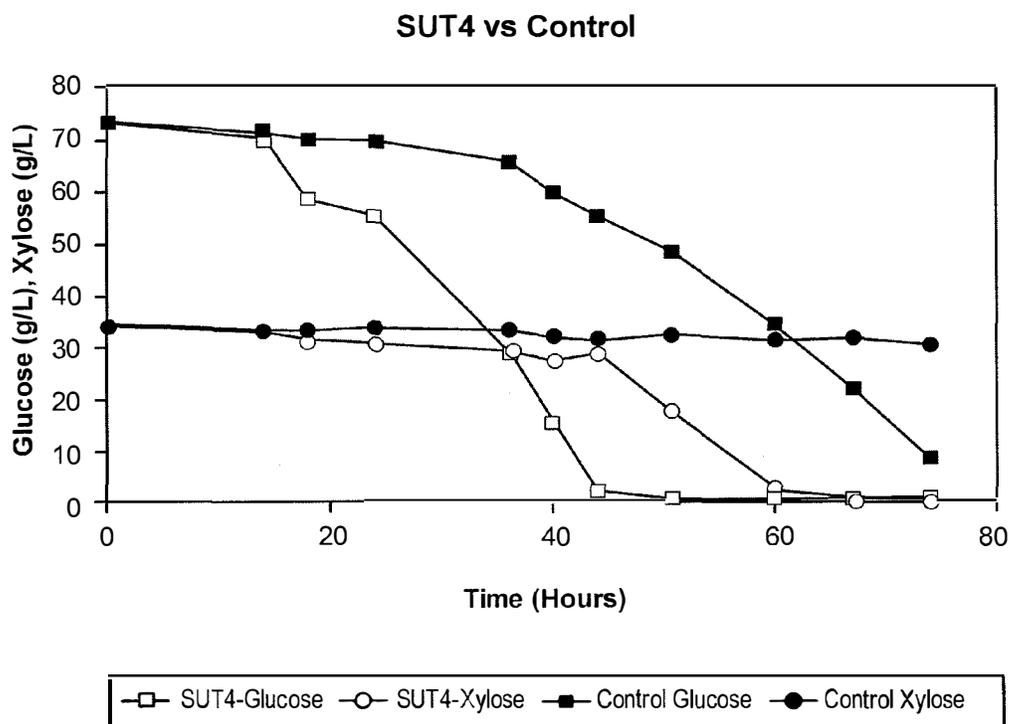


FIG. 5

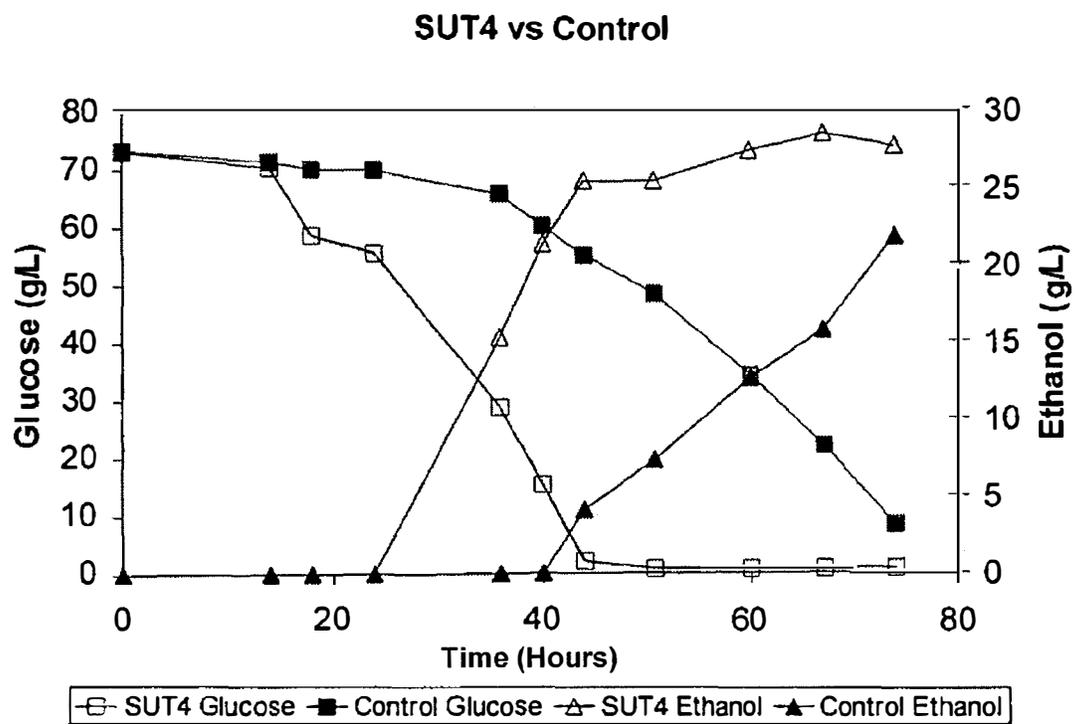


FIG. 6

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SUGAR TRANSPORT SEQUENCES, YEAST STRAINS HAVING IMPROVED SUGAR UPTAKE, AND METHODS OF USE

CROSS-REFERENCE TO RELATED APPLICATIONS

This application is a continuation of U.S. patent application Ser. No. 12/484,714, filed Jun. 15, 2009, now U.S. Pat. No. 8,105,811, which claims priority to U.S. Provisional Patent Application No. 61/061,417 filed Jun. 13, 2008, the contents of which are incorporated by reference in their entirety.

REFERENCE TO A "SEQUENCE LISTING," A TABLE, OR A COMPUTER PROGRAM LISTING APPENDIX SUBMITTED AS AN ASCII TEXT FILE

The Sequence Listing written in file -23-2.TXT, created on Dec. 28, 2011, 24,576 bytes, machine format IBM-PC, MS-Windows operating system, is hereby incorporated by reference in its entirety for all purposes.

STATEMENT REGARDING FEDERALLY SPONSORED RESEARCH OR DEVELOPMENT

This invention is jointly owned between WARF and the USDA and was made with United States government support awarded by the following agencies:

USDA Grant Number 2006-35504-17436.

The government has certain rights in this invention.

BACKGROUND OF THE INVENTION

Within the United States, ongoing research is directed toward developing alternative energy sources to reduce our dependence on foreign oil and nonrenewable energy. The use of ethanol as a fuel has become increasingly prevalent in recent years. Currently, corn is the primary carbon source used in ethanol production. The use of corn in ethanol production is not economically sustainable and, arguably, may result in increased food costs.

In order to meet the increased demand for ethanol, it will be necessary to ferment sugars from other biomass, such as agricultural wastes, corn hulls, corncobs, cellulosic materials, pulping wastes, fast-growing hardwood species, and the like. Biomass from most of these sources contains large amounts of xylose, constituting about 20 to 25% of the total dry weight. Because agricultural residues have a high xylose content, the potential economic and ecologic benefits of converting xylose in these renewable materials to ethanol are significant.

In biomass conversion, microorganisms serve as biocatalysts to convert cellulosic materials into usable end products such as ethanol. Efficient biomass conversion in large-scale industrial applications requires a microorganism that can tolerate high sugar and ethanol concentrations, and which is able to ferment multiple sugars simultaneously.

The pentose D-xylose is significantly more difficult to ferment than D-glucose. Bacteria can ferment pentoses to ethanol and other co-products, and bacteria with improved ethanol production from pentose sugars have been genetically engineered. However, these bacteria are sensitive to low pH and high concentrations of ethanol, their use in fermentations is associated with co-product formation, and the level of

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ethanol produced remains too low to make the use of these bacteria in large-scale ethanol production economically feasible.

In general, industrial producers of ethanol strongly favor using yeast as biocatalysts, because yeast fermentations are relatively resistant to contamination, are relatively insensitive to low pH and ethanol, and are easier to handle in large-scale processing. Many different yeast species use xylose respiratively, but only a few species use xylose fermentatively. Fermentation of xylose to ethanol by wild type xylose-fermenting yeast species occurs slowly and results in low yields, relative to fermentation rates and ethanol yields that are obtained with conventional yeasts in glucose fermentations. In order to improve the cost effectiveness of xylose fermentation, it is necessary to increase the rate of fermentation and the ethanol yields obtained.

The most commonly used yeast in industrial applications is *Saccharomyces cerevisiae*. Although *S. cerevisiae* is unable to grow on or ferment xylose, homogenates of *S. cerevisiae* can readily ferment D-ribulose-5-phosphate to ethanol, and can convert D-xylulose-5-phosphate to a lesser extent. *S. cerevisiae* metabolically engineered to overproduce D-xylose reductase (XYL1), xylitol dehydrogenase (XYL2) and D-xy-lulokinase (XYL3 or XKS1) or some forms of xylose isomerase (xylA) along with XYL3 or XKS1 can metabolize xylose to ethanol.

Pichia stipitis is a yeast species that in its native state is able to ferment xylose to produce ethanol. In *P. stipitis*, fermentative and respirative metabolism co-exist to support cell growth and the conversion of sugar to ethanol.

There is a need in the art for yeast strains having improved ability to convert sugars to ethanol.

BRIEF SUMMARY OF THE INVENTION

In one aspect, the present invention provides a nucleic acid construct comprising a coding sequence operably linked to a promoter not natively associated with the coding sequence. The coding sequence encodes a polypeptide having at least 95% amino acid identity to SEQ ID NO:2, SEQ ID NO:4, or SEQ ID NO:6.

Other aspects of the invention provide yeast strains comprising the nucleic acid construct, and methods of producing ethanol or xylitol by growing a yeast strain of the invention in xylose-containing medium.

BRIEF DESCRIPTION OF THE SEVERAL VIEWS OF THE DRAWINGS

FIG. 1 shows the alignment of PsSUT2, PsSUT3, and PsSUT4.

FIG. 2 shows growth, measured as OD₆₀₀, of *S. cerevisiae* expressing XUT1 or SUT4 on xylose as a function of time.

FIG. 3 shows growth, measured as OD₆₀₀, of *S. cerevisiae* expressing XUT1 or SUT4 on glucose as a function of time.

FIG. 4 shows uptake of ¹⁴C-glucose or ¹⁴C-xylose, measured as DPM, by *S. cerevisiae* expressing XUT1 or SUT4 as a function of time.

FIG. 5 shows consumption of glucose and xylose by *Pichia stipitis* overexpressing SUT 4 as a function of time.

FIG. 6 shows glucose consumption and ethanol production by *Pichia stipitis* overexpressing SUT4 as a function of time.

DETAILED DESCRIPTION OF THE INVENTION

One approach to increasing the efficiency of yeast-catalyzed conversion of sugars to ethanol or to other products

such as xylitol is to enhance uptake of sugars by the yeast. Cellulosic and hemicellulosic materials used in fermentation reactions are generally subjected to pretreatments and enzymatic processes that produce a mixture of several sugars, including glucose, xylose, mannose, galactose, and arabinose.

Saccharomyces cerevisiae and other hexose-fermenting yeasts have sugar transporters that facilitate uptake of hexoses. Some hexose sugar transporters also promote uptake of pentoses such as xylose, but at a much lower rate and with much lower affinity. Therefore, xylose uptake and utilization is impaired until glucose is consumed. When the substrate includes large initial concentrations of glucose, ethanol production from fermentation of glucose may result in inhibitory concentrations of ethanol before xylose uptake begins.

In order to promote xylose utilization by yeast in fermentation of mixed hemicellulosic sugars, sugar transport proteins having high affinity or specificity for xylose were identified and used to develop yeast engineered to have improved xylose uptake, as described below in the Examples.

The genome of the xylose-fermenting yeast *Pichia stipitis* was examined for putative sugar transporters based on similarity to transporters that facilitate uptake of hexoses, lactose, maltose, phosphate, urea, and other compounds. Eight putative xylose transporters (XUT1-7 and SUT4) were chosen based on their implied structures and sequence similarities to other reported sugar transporters. Genome array expression studies were used to evaluate whether the sugar transporters showed induced expression in cells grown on xylose. Genome array expression results were confirmed using quantitative PCR of transcripts from cells grown on xylose.

Sequences encoding polypeptides corresponding to those encoded by two genes (XUT1 and SUT4) that showed increased expression in cells grown on xylose-containing media under oxygen limited conditions were expressed in a host strain *Saccharomyces cerevisiae* JYB3011, which lacks all hexose-transport genes and expresses *P. stipitis* XYL1, XYL2, and XYL3, which encode xylose reductase (XR), xylitol dehydrogenase (XDH), and xylulokinase (XK), respectively. Expression of either XUT1 or SUT4 allowed growth of the *S. cerevisiae* strain on xylose. Both XUT1 and SUT4 allowed increased xylose uptake. Cells carrying these sequences were tested for their relative affinities in taking up ¹⁴C-labeled glucose or ¹⁴C-labeled xylose. Xut1 was shown to have a higher affinity for xylose, and Sut4 a higher affinity for glucose. A transformant carrying a partial sequence of the XUT1 gene, which encodes the C-terminal portion of the protein, beginning with amino acid residue 154 of SEQ ID NO:2, was shown to bind xylose and glucose but did not accumulate these sugars, thereby demonstrating that at least some portion of the amino terminus of the protein is needed for sugar transport.

Thus, expression of any of XUT1, XUT1 partial, and SUT4 in *Saccharomyces cerevisiae* allows growth on xylose, and in the case of XUT1 and SUT4, uptake of xylose. It is envisioned that expression of any of XUT1, XUT1 partial, and SUT4 or sequences similar to XUT1, XUT1 partial, and SUT4 in other yeast species will also allow growth on xylose and/or uptake of xylose.

The sequence of the deduced amino acid sequence of the putative proteins encoded by the *Pichia stipitis* XUT1 and SUT4 genes are provided in SEQ ID NO:2 and SEQ ID NO:4, respectively. The sequences of the XUT1 and SUT4 coding sequences prepared from *Pichia stipitis* are shown in SEQ ID NO:1 and SEQ ID NO:3, respectively.

The *P. stipitis* glucose/xylose facilitators PsSut1-4 have a high degree of sequence identity, with PsSut1 having 78%

amino acid identity with PsSut2, 3, and 4. PsSut2 differs from PsSut3 by one amino acid (L279V). PsSut4 differs from PsSut2 by just one amino acid (A544T) and from PsSUT3 by two amino acids (L279V and A544T). While all four facilitators have similar affinities for glucose, their affinities for xylose were shown to differ significantly (See Table 3). Notably, PsSut4 has a higher specificity ratio (V.sub.max/K.sub.m) than the other Sut facilitators for glucose and xylose, and a higher affinity for xylose than any of the other facilitators.

It is envisioned that changes to the polypeptides could be made without affecting their activities, and such variations are within the scope of the invention. For example, polypeptides have insertions, deletions or substitutions of amino acids relative to SEQ ID NO:2 and SEQ ID NO:4 are within the scope of the invention, provided that the polypeptide has at least 80% amino acid identity to SEQ ID NO:2 and SEQ ID NO:4. Suitably, a polypeptide according to the invention has at least 85%, 90%, or 95% amino acid identity to SEQ ID NO:2 or SEQ ID NO:4. It is further envisioned that polypeptides according to the invention have at least 80%, 85%, 90%, 95%, 96%, 97%, 98%, 99%, or 100% amino acid identity to SEQ ID NO:4, and have a threonine residue or a serine residue at a position corresponding to amino acid 544 of SEQ ID NO:4. Preferably, polypeptides according to the invention retain their activity, i.e., retain the ability to enhance uptake of glucose and/or xylose. Such activity may be assayed by any suitable means, including, for example, those described in the examples.

It is well understood among those of ordinary skill in the art that certain changes in nucleic acid sequence make little or no difference to the overall function of the protein or peptide encoded by the sequence. Due to the degeneracy of the genetic code, particularly in the third position of codons, changes in the nucleic acid sequence may not result in a different amino acid being specified by that codon. Changes that result in an amino acid substitution may have little or no effect on the three dimensional structure or function of the encoded protein or peptide. Conservative substitutions are even less likely to result in a lost of function. In addition, changes that result in insertions or deletions of amino acids may also be acceptable.

Alignment of the native sequence encoding PsSut4 with those encoding PsSut2 and PsSut3 shows that PsSut4 differs from the coding sequences for PsSut2 and PsSut3 at more than 30 bases (FIG. 1), with only two differences resulting in a change in the amino acid specified. Suitably, polynucleotides according to the invention include one or more of the divergent sequences of PsSUT4 shown in FIG. 1, in any combination. Suitably, the polynucleotides include anywhere from one to all of the divergent sequences of PsSUT4 shown in FIG. 1.

It is expected that expression of more than one of the proteins encoded by the polynucleotides of the invention, as exemplified by SEQ ID NO:2, SEQ ID NO:4, and SEQ ID NO:6, could be obtained in a transgenic yeast cell. Expression of two or more polynucleotides of the invention may afford additional advantages in terms of fermentation rate or sugar uptake. For example, one could create a yeast strain that expresses a polynucleotide encoding a polypeptide having at least 80%, at least 85%, at least 90%, or at least 95% amino acid identity to SEQ ID NO:2 together with a polynucleotide encoding a polypeptide having at least 80%, at least 85%, at least 90%, or at least 95% amino acid identity to SEQ ID NO:4.

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Additionally, the polynucleotides of the invention may further include a sequence encoding a selectable marker that would allow selection of yeast expressing the polynucleotide.

In the examples, expression of SUT4 under the control of the promoter from *P. stipitis* fatty acid synthase 2 (FAS2 promoter), which is induced by xylose and under oxygen limiting conditions, was obtained in *S. cerevisiae* yJML123 and in *P. stipitis* UC7. The yJML123+SUT4 was able to use both glucose and xylose at a faster rate than the control strain. Neither strain was able to use all of the xylose. The yJML123+SUT4 strain, which has reduced expression of xylitol reductase, produced a higher yield of xylitol (18.45 g/L) than the control strain (15.51 g/L). Further, the yJML123+SUT4 strain produced a higher yield of xylitol faster than the control.

The *P. stipitis* UC7+SUT4 strain was grown under oxygen limiting conditions (50 mL of culture with a starting OD₆₀₀ of 10 in a 125 mL flask aerated at speed of 100 rpm at 30° C.). The UC7+SUT4 strain was found to use xylose at a faster than the UC7 control; neither strain used all of the xylose. The xylitol yield of UC7+SUT4 strain (14.98 g/L) was similar to that of the UC7 control (15.18 g/L). The UC7+SUT4 strain had a higher ethanol specific productivity.

In another example, UC7+SUT4 strain was grown in glucose (7% w/w) and xylose (3% w/w) in a three liter bioreactor. Those results indicated that the UC7+SUT4 grew at a faster rate than did the UC7 control. The UC7+SUT4 strain had a higher biomass yield than the control. Additionally, the ethanol yield was higher for the UC7+SUT4 strain. The UC7+SUT4 strain consumed both glucose and xylose at a higher rate and consumed all of the sugars, whereas the control consumed only 88% of the glucose and did not appear to consume xylose. Finally, the specific rate of ethanol production on a dry cell weight basis was increased by more than three fold.

In the examples, strains were developed by causing nucleic acid of the invention to be introduced into a chromosome of the host yeast strain. This can be accomplished by any suitable means. However, it is envisioned that one could obtain increased expression of the nucleic acid constructs of the invention using an extrachromosomal genetic element, by integrating additional copies, by increasing promoter strength, or by increasing the efficiency of translation through codon optimization, all methods known to one of skill in the art.

It is envisioned that in addition to the particular strains that were used, any strain of *S. cerevisiae* or *P. stipitis* could be used in the practice of the invention. For example, in addition to *P. stipitis* UC7, one could advantageously obtain expression of the polynucleotides of the invention in *P. stipitis* strains having reduced respiration capacity, such as petite mutants or *P. stipitis* Shi21 (NRRL Y-21971). Similarly, one could advantageously obtain expression of the polynucleotides of the invention in a PHO-13 mutant of *S. cerevisiae* expressing *P. stipitis* XYL1, XYL2, and XYL3. Such a strain was developed, as described in the examples, for use in developing yeast strains according to the invention. Yeast strains may suitably be modified to include exogenous sequences expressing Xut1 and Sut4 on the same or different nucleic acid. The yeast strains may be modified to include additional advantageous features, as would be appreciated by one skilled in the art.

In addition to *S. cerevisiae* and *P. stipitis*, it is envisioned that other yeast species could be used to obtain yeast strains according to the invention for use in the methods of the invention. Other suitable yeast species include, without limitation, *Candida boidinii*, *Enteroramus dimorphus*, *Candida*

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jeffriesii, *Debaryomyces hanseii*, *Candida Guillermundii*, *Candida shehatae*, *Brettanomyces naardensis*, *Candida guillermundii*, *Candida lyxosophilia*, *Candida intermedia*, *Candida tenuis*, *Hansenula polymorpha*, *Kluyveromyces marxianus*, *Kluyveromyces lactis*, *Kluyveromyces fragilis*, *Kluyveromyces thermotolerans*, *Pachysolen tannophilus*, *Pichia segobiensis*, *Spathaspora passalidarum* and Pass 1 isolates.

In another aspect, the present invention provides a method of fermenting xylose in a xylose-containing material to produce ethanol using the yeast of the invention as a biocatalyst. Another aspect of the present invention provides a method of fermenting xylose in a xylose-containing material to produce xylitol using the yeast of the invention as a biocatalyst. In this embodiment, the yeast preferably has reduced xylitol dehydrogenase activity such that xylitol is accumulated. Preferably, the yeast is recovered after the xylose in the medium is fermented to ethanol and used in subsequent fermentations.

The constructs of the invention comprise a coding sequence operably connected to a promoter. Preferably, the promoter is a constitutive promoter functional in yeast, or an inducible promoter that is induced under conditions favorable to uptake of sugars or to permit fermentation. Inducible promoters may include, for example, a promoter that is enhanced in response to particular sugars, or in response to oxygen limited conditions, such as the FAS2 promoter used in the examples. Examples of other suitable promoters include promoters associated with genes encoding *P. stipitis* proteins which are induced in response to xylose under oxygen limiting conditions, including, but not limited to, myo-inositol 2-dehydrogenase (MOR1), aminotransferase (YOD1), guanine deaminase (GAH1). These proteins correspond to protein identification numbers 64256, 35479, and 36448 on the Joint Genome Institute *Pichia stipitis* web site: genome.jgi-psf.org/Picst3/Picst3.home.html.

Oxygen limiting conditions include conditions that favor fermentation. Such conditions, which are neither strictly anaerobic nor fully aerobic, can be achieved, for example, by growing liquid cultures with reduced aeration, i.e., by reducing shaking, by increasing the ratio of the culture volume to flask volume, by inoculating a culture medium with a number of yeast effective to provide a sufficiently concentrated initial culture to reduce oxygen availability, e.g., to provide an initial cell density of 1.0 g/l dry wt of cells.

By "xylose-containing material," it is meant any medium comprising xylose, whether liquid or solid. Suitable xylose-containing materials include, but are not limited to, hydrolysates of polysaccharide or lignocellulosic biomass such as corn hulls, wood, paper, agricultural by-products, and the like.

By a "hydrolysate" as used herein, it is meant a polysaccharide that has been depolymerized through the addition of water to form mono and oligosaccharides. Hydrolysates may be produced by enzymatic or acid hydrolysis of the polysaccharide-containing material, by a combination of enzymatic and acid hydrolysis, or by an other suitable means.

Preferably, the yeast strain is able to grow under conditions similar to those found in industrial sources of xylose. The method of the present invention would be most economical when the xylose-containing material can be inoculated with the mutant yeast without excessive manipulation. By way of example, the pulping industry generates large amounts of cellulosic waste. Saccharification of the cellulose by acid hydrolysis yields hexoses and pentoses that can be used in fermentation reactions. However, the hydrolysate or sulfite liquor contains high concentrations of sulfite and phenolic inhibitors naturally present in the wood which inhibit or pre-

vent the growth of most organisms. Serially subculturing yeast selects for strains that are better able to grow in the presence of sulfite or phenolic inhibitors.

It is expected that yeast strains of the present invention may be further manipulated to achieve other desirable characteristics, or even higher specific ethanol yields. For example, selection of mutant yeast strains by serially cultivating the mutant yeast strains of the present invention on medium containing hydrolysate may result in improved yeast with enhanced fermentation rates.

The following non limiting examples are intended to be purely illustrative.

Examples

Characterization of Putative Xylose Transporters of *Pichia stipitis*

Yeast strains, plasmids, and growth conditions. The yeast strains and the plasmids used in this study are listed in Table 1. *S. cerevisiae* EB.Y.VW4000 strain (MATa Dhxt1-17 Δgal2 Δstl1 Δagt1 Δmph2 Δmph3 leu2-3,112 ura3-52 trp1-289 his3-Δ1 MAL2-8c SUC2) (9) was kindly provided by E. Boles at Heinrich-Heine-Universität and was used for transformation of all the constructed plasmids. Three xylose transporter candidates, XUT1, XUT1 partial (corresponding to the sequence encoding the C-terminal region of the protein beginning with amino acid residue 154 of SEQ ID NO:2), and SUT4, were amplified and fused with the TDH3 promoter by PCR. The coding sequence of SUT4 includes a CUG codon, which in *P. stipitis*, specifies the serine residue of amino residue 410 of SEQ ID NO:4. This codon, which in most organisms specifies a leucine, was modified for expression in *S. cerevisiae* using standard methods so as to specify a serine residue. Each candidate had its own terminator. These fragments were then inserted into pRS316 after sequence confirmation (Sikorski & Hieter (1989) Genetics 122, 19-27). Genetic manipulation and cloning of DNA were performed using methods known in the art. *Escherichia coli* DH5α.F' was used as a host for plasmid preparation.

Yeast transformation was performed as previously described (Gietz & Woods (2002) Methods Enzymol 350, 87-96). Yeast transformants were selected by TRP1 and URA3 selectable markers and cultivated on yeast synthetic complete dropout (TRP-URA-) medium with carbon source (Kaiser et al. & Cold Spring Harbor Laboratory. (1994) Methods in yeast genetics: a Cold Spring Harbor Laboratory course manual (Cold Spring Harbor Laboratory Press, Cold Spring Harbor, N.Y.)). Yeast cells were cultured in a 125-ml flask with 50 ml medium at 30° C. and 200 rpm.

Quantitative PCR (qPCR)

Total RNA was prepared from each transformant and 5 μg of total RNA was used for cDNA construction with random oligonucleotides and the Reverse Transcription System Kit (Promega, Madison, Wis.). Reverse Transcription-PCR primers were designed using Primer Express software (Applied Biosystems). RT-PCR was performed with SYBR green PCR master mix (Applied Biosystems) and an ABI PRISM 7000 sequence detection system (Applied Biosystems). PCR conditions were as recommended by the manufacturer except that one-half of the reaction volume was used. Actin was used to normalize the relative copy numbers of each gene. All reactions were performed in triplicate.

TABLE 1

List of yeast strains and plasmids		
Strain or plasmid	Description	Source or reference
Strains		
<i>S. cerevisiae</i> EB.Y.VW4000	MATa Dhxt1-17 Δgal2 Δstl1 Δagt1 Δmph2 Δmph3 leu2-3, 112 ura3-52 trp1-289 his3-Δ1 MAL2-8c SUC2	Wieczorke
<i>S. cerevisiae</i> JYB3005	<i>S. cerevisiae</i> EB.Y.VW4000 (pRS314 and pRS316)	This study
<i>S. cerevisiae</i> JYB3011	<i>S. cerevisiae</i> EB.Y.VW4000 (pRS314-X123 and pRS316)	This study
<i>S. cerevisiae</i> JYB3110	<i>S. cerevisiae</i> EB.Y.VW4000 (pRS314-X123 and pRS316-XUT1partial)	This study
<i>S. cerevisiae</i> JYB3210	<i>S. cerevisiae</i> EB.Y.VW4000 (pRS314-X123 and pRS316-XUT1)	This study
<i>S. cerevisiae</i> JYB3310	<i>S. cerevisiae</i> EB.Y.VW4000 (pRS314-X123 and pRS316-SUT4)	This study
Plasmids		
pRS314	TRP1 CEN/ARS	Sikorski
pRS316	URA3 CEN/ARS	Sikorski
pRS314-X123	TRP1 CEN/ARS TDH3 _p -PsXYL1-TDH3 _T TDH3 _p -PsXYL2-TDH3 _T TDH3 _p -PsXYL3-TDH3 _T	Ni
pRS316-XUT1partial	URA3 CEN/ARS TDH3 _p -PsXUT1	This study
pRS316-XUT1	URA3 CEN/ARS TDH3 _p -PsXUT1	This study
pRS316-SUT4	URA3 CEN/ARS TDH3 _p -PsSUT4	This study

Sugar-Transport Assay.

Initial D-[14C]xylose and D-[14C]glucose (Amersham) uptake rates were measured at 30° C., pH 5.0 as previously described (Spencer-Martins, I. a. c. U., N. (1985) Biochim. Biophys. Acta 812, 168-172). Cells were harvested in the mid-exponential phase (A₆₀₀ nm 0.8-1.0; ≈0.1-0.125 g/l dry wt) by centrifugation, washed twice with ice-cold water and resuspended to a cell concentration of 5-10 mg (dry weight)/ml. Kinetic parameters were determined using Lineweaver-Burk plots. All measurements were conducted at least three times, and the data reported are the average values.

Transcriptional Analysis of Eight Xylose Transporter Candidates.

Eight putative xylose transporters (XUT1-7 and SUT4) were identified based on genomic analysis of the genome sequence of *Pichia stipitis* (Jeffries et al. (2007) Nat Biotechnol 25, 319-26) and selected for further analysis. Of these eight putative transporters, XUT1, and SUT4 showed increased expression in *Pichia stipitis* grown in xylose-containing media. These two genes were introduced into the host strain *S. cerevisiae* JYB3011 to evaluate the ability of the putative xylose transporters to promote xylose uptake. This host strain was originated from *S. cerevisiae* EB.Y.VW4000 (Wieczorke et al. (1999) FEBS Lett 464, 123-8) and engineered to express PsXYL1, PsXYL2, and PsXYL3 by introducing pRS314-X123 (Ni et al. (2007) Appl Environ Microbiol 73, 2061-6) (Table 1).

Kinetic Characteristics of Sugar Uptake.

XUT1 partial, XUT1 and SUT4 were expressed in the strains *S. cerevisiae* JYB3110, JYB3210, and JYB3310, respectively (Table 1). The expression of each transporter in its respective strain was confirmed by qPCR (data not shown). These three strains could grow in xylose-containing media (FIG. 2) and in glucose-containing media (FIG. 3). Xylose and glucose transport capacity of each candidate was tested using a high concentration (50 mM) of labeled sugar (Table 2). The strains containing Xut1 or Sut4 showed increased

xylose uptake activities, with the strain expressing Xut1 having a higher uptake rate than that of the strain expressing Sut4 (FIG. 4). Both JYB 3210 and JYB3310 showed glucose-transporting activity, with Sut4 conferring a higher glucose uptake velocity than Xut1 (FIG. 4).

Kinetic analysis demonstrated that the affinity of Xut1 for xylose is 3-fold higher than its affinity for glucose (FIG. 4 and Table 3). This is the first reported transporter having a higher affinity for xylose. V_{max} was also higher with xylose than glucose. Although Sut4 exhibits xylose uptake activity, its affinity for glucose is 12 times higher than its affinity for xylose (Table 3). Interestingly, Sut4 has higher V_{max} with xylose than glucose. Notably, Sut4 has a higher specificity ratio (V_{max}/K_m) for both xylose and glucose than any of the other four glucose/xylose facilitators found in *Pichia stipitis* (Table 3).

Table 3 compares K_m values of the newly discovered xylose transporters to previously characterized xylose trans-

ported to be a high affinity xylose-proton symporter, and SUT4 appears to employ facilitated diffusion.

TABLE 2

Initial uptake rates of labeled sugar (50 mM) of each putative xylose transporter.		
Strain	Substrate	V [†]
JYB3011 (Control)	Glucose	5
	Xylose	0
JYB3110 (XUT1 partial)	Glucose	7
	Xylose	0
JYB3210 (XUT1)	Glucose	31.8
	Xylose	57.8
JYB3310 (SUT4)	Glucose	52.6
	Xylose	23.1

[†]Velocity of uptake (micromoles per minutes per milligram [dry weight])

TABLE 3

Kinetic parameters of yeast glucose/xylose transporters							
Transporters	K _m (mM)		V _{max} (nmol min ⁻¹ mg dw ⁻¹)		Specificity V _{max} /K _m		Reference
	Glucose	Xylose	Glucose	Xylose	Glucose	Xylose	
PsSut1	1.5 ± 0.1	145.0 ± 1.0	45 ± 1.0	132 ± 1.0	30.0	0.9	(4)
PsSut3	0.8 ± 0.1	103 ± 3.0	3.7 ± 0.1	87 ± 2.0	4.6	0.8	(4)
PsSut2	1.1 ± 0.1	49.0 ± 1.0	3.3 ± 0.1	28 ± 4.0	3.3	0.6	(4)
PsSut4	1.3 ± 0.1	16.6 ± 0.3	105 ± 4.2	122 ± 2.4	80.8	7.4	This study
CiGxf1	2.0 ± 0.6	48.7 ± 6.5	≈163 [†]	≈25 [†]	81.5	0.5	(1)
ScHxt1	^a 107 ± 49	^b 880 ± 8	^a 50.9 ± 3.7	^b 750 ± 94	0.5	0.8	^a (2) ^b (3)
ScHxt2	^a 2.9 ± 0.3	^b 260 ± 130	^a 15.6 ± 0.9	^b 340 ± 10	5.4	1.3	^a (2) ^b (3)
ScHxt4	^a 6.2 ± 0.5	^b 170 ± 120	^a 12.0 ± 0.9	^b 190 ± 23	1.9	1.1	^a (2) ^b (3)
ScHxt7	^a 1.3 ± 0.3	^b 130 ± 9	^a 11.7 ± 0.3	^b 110 ± 7	90.0	0.8	^a (2) ^b (3)
CiGxs1	0.012 ± 0.004	0.4 ± 0.1	4.3 ± .33	6.5 ± 1.5	358.3	16.2	(1)
PsXut1	0.91 ± 0.01	0.46 ± 0.02	80 ± 1.0	116 ± 5.8	87.91	252.2	This study

[†]Calculated from FIG. 2 of (1)

1. Leandro, M. J., P. Goncalves, and I. Spencer-Martins. 2006. Two glucose/xylose transporter genes from the yeast *Candida intermedia*: first molecular characterization of a yeast xylose-H⁺ symporter. *The Biochemical journal* 395:543-549.
2. Maier, A., B. Volker, E. Boles, and G. F. Fuhrmann. 2002. Characterisation of glucose transport in *Saccharomyces cerevisiae* with plasma membrane vesicles (countertransport) and intact cells (initial uptake) with single Hxt1, Hxt2, Hxt3, Hxt4, Hxt6, Hxt7 or Gal2 transporters. *Fems Yeast Research* 2:539-550.
3. Saloheimo, A., J. Rauta, V. Stasyk, A. A. Sibiry, M. Penttila, and L. Ruohonen. 2007. Xylose transport studies with xylose-utilizing *Saccharomyces cerevisiae* strains expressing heterologous and homologous permeases. *Applied Microbiology And Biotechnology* 74:1041-1052.
4. Weierstall, T., C. P. Hollenberg, and E. Boles. 1999. Cloning and characterization of three genes (SUT1-3) encoding glucose transporters of the yeast *Pichia stipitis*. *Molecular Microbiology* 31:871-883.

porters. All four of the PsSut glucose/xylose facilitators show similar affinities for glucose, but their affinities for xylose differ significantly. Notably, PsSut4 shows a higher specificity ratio (V_{max}/K_m) for glucose and xylose than the specificity ratios of PsSut1-3. Interestingly, PsSut1 has about 78% amino acid identity with PsSut2, 3 and 4, whereas PsSut2 and PsSut4 differ from PsSut3 by one or two amino acids each (Sut2: L279V; Sut4: L279V, A544T). Despite the high degree of sequence identity, PsSut1-4 have distinct properties. PsSut1 and PsSut3 can mediate fructose transport, whereas PsSut2 cannot, and only PsSut3 can mediate galactose transport (Weierstall et al. (1999) *Molecular Microbiol.* 31:871-883). Notably, among PsSut1-4, PsSut4 has the highest affinity and the second highest V_{max} for xylose. The affinity of PsSut4 for xylose is about three times higher than that of PsSut2, and its V_{max} for xylose uptake is about 4.4 times higher than the V_{max} of Sut2.

Energy requirements for sugar transport were evaluated by including the metabolic uncoupler sodium azide. Xut1 mediated uptake of sugar was reduced by 96% in the presence of sodium azide, whereas Sut4 mediated sugar uptake was reduced by about 50% (Table 4). Consequently, Xut1 is

TABLE 4

	Effect of sodium azide on glucose and xylose uptake [†] .			
	Glucose		Xylose	
	Control	NaN ₃ (50 mM)	Control	NaN ₃ (50 mM)
JYB3110 (SUT4)	41	0.32	90	3.19
JYB3310 (XUT1)	40	25	13.4	9.2

[†]Velocity of uptake (nmol min⁻¹ mg⁻¹ [dry weight])

Effect of Expression of Sut4 in *P. stipitis*

Construction of Plasmids and Transformation of Yeast Strains.

A plasmid was constructed to contain the *P. stipitis* coding sequence for the SUT4 putative transporter under the control of the *P. stipitis* fatty acid synthase subunit alpha (Fas2) promoter (FAS2p_SUT4_SUT4t_LoxP_URA3_LoxP plasmid). Roughly 100 ug of the FAS2p_SUT4_SUT4t_LoxP_URA3_LoxP plasmid was linearized using Hind III and ApaI, ethanol precipitated and resuspended in water to create a fragment that would integrate

directly into the *Pichia* genome. The digested construct was then used in a LiAc protocol and transformed into either the yJML123 (Gietz & Woods (2002) Methods Enzymol 350, 87-96) or UC7 *Pichia stipitis* strains. The yJML123 strain has a deletion in the XYL2 gene, which encodes the enzyme that catalyzes the conversion of xylitol to xylulose, allowing us to create *Pichia* strains that will begin to accumulate xylitol.

Transformants were selected for via growth on ScD-Ura plates, which contain 0.62 g/L CSM-Leu-Trp-Ura (Bio 101 Systems) and dextrose (2%). Colonies were then picked and grown overnight in ScD-Ura liquid media. Genomic DNA was then extracted and evaluated by PCR to confirm integration of the construct. As a control for these strains, a plasmid containing only the LoxP_Ura3_LoxP cassette was also integrated into the yJML123 and UC7 strains (yJML123 and UC7 controls).

Fermentation of yJML123+SUT4

Starter cultures were established by inoculating a single colony of yJML123+SUT4 strain or yJML123 control from recently streaked plate into 25 mL fermentation media and grown overnight. The following morning, triplicate flask cultures of each transformant were started at an OD₆₀₀ of 0.5 (≈ 0.06 g/l dry wt of cells). The transformants were confirmed using PCR genotyping. The fermentation was run under aerobic conditions using an agitation speed of 200 rpm at 30° C. in 25 mL of media in a 125 mL flask in fermentation medium (Yeast Nitrogen Base (1.7 g/L), urea (2.4 g/L), peptone (3.6 g/L), with xylose (40 g/L) and glucose (17 g/L) as carbon sources. Samples were collected for analysis roughly every 8 hours for the first 48 hours and then once every 24 hours.

The yJML123+SUT4 strain was able to use both glucose and xylose at a faster rate than the control strain. Neither strain was able to use all of the xylose. The yJML123+SUT4 strain produced a higher yield of xylitol (18.45 g/L) than the control strain (15.51 g/L). Further, the yJML123+SUT4 strain produced a higher yield of xylitol faster than the control.

Fermentation of UC7+SUT4.

Starter cultures were established by inoculating a single colony of SUT4+UC7 strain or the UC7 control from recently streaked plates into 50 mL of fermentation media and grown overnight. The following morning, triplicate flasks of each transformant were started at an OD₆₀₀ of 0.5 (≈ 0.06 g/l dry wt of cells). The transformants were confirmed using PCR genotyping. The fermentation was run in fermentation medium (Yeast Nitrogen Base (1.7 g/L), urea (2.4 g/L), peptone (3.6 g/L), and xylose (40 g/L), under low oxygen conditions using an agitation speed of 100 rpm at 30° C. in 50 mL of media in a 125 mL flask. The starting OD₆₀₀ was 10 (≈ 1.25 g/l dry wt of cells). Samples were collected for analysis roughly every 12 hours for the first 60 hours and then once every 24 hours.

Fermentation of UC7+SUT4.

The UC7+SUT4 strain was found to use xylose at a faster rate than the UC7 control; neither strain used all of the xylose. The xylitol yield of UC7+SUT4 strain was similar to that of the UC7 control. The UC7+SUT4 strain had a higher ethanol specific productivity than that of the UC7 control.

Evaluation of Expression of SUT4 in *P. stipitis* in Bioreactor Strain Descriptions.

The UC7+SUT4 strain described above ("SUT4") and the UC7+URA3 cassette strain ("control") described above were evaluated in a three liter bioreactor scale-up as described below.

3 L Bioreactor Scale-up of the SUT4 Transporter Overexpression Strain.

Low oxygen bioreactor trials were used to compare two strains in a larger scale and under more controlled conditions than shake flask trials.

Bioreactor Cultivations.

A defined minimal medium containing trace metal elements and vitamins was used in all bioreactor cultivations. This medium was modified based on that described by Verduyn et al. (Verduyn et al. (1992) Yeast 8:501-517) to include urea as a nitrogen source. It had the following composition: 2.4 g urea L⁻¹; 3 g KH₂PO₄ L⁻¹, 0.5 g MgSO₄ 7 H₂O L⁻¹; 1 ml trace element solution L⁻¹; 1 ml vitamin solution L⁻¹; and 0.05 ml antifoam 289 (Sigma A-8436) L⁻¹ (Jeffries & Jin (2000) Adv Appl Microbiol 47, 221-68). For these cultivations, a starting concentration of 70 g L⁻¹ glucose and 30 g L⁻¹ xylose was used.

Cultivations were performed in 3 L New Brunswick Scientific BioFlo 110 Bioreactors with working volumes of 2 liters. All cultivations were performed at 30° C. Agitation was set at 500 rpm and the pH was controlled at pH 5.0 by the addition of 5 N KOH and 5 N H₂SO₄. Aeration was controlled at a rate of 0.5 vvm which corresponded to a rate of 1 L min⁻¹. Input gas was mixed using a gas proportioner to include 90% pure nitrogen and 10% air, for a final oxygen concentration of approximately 2%. Bioreactors were inoculated to an OD of approximately 1 and their progress followed for 74 hours.

During cultivation, the SUT4 strain appeared to grow faster than the control. Calculated biomass rates and yields demonstrated that SUT4 had a slightly improved growth rate (~1.2 fold) (0.062 vs. 0.0514 h⁻¹) and biomass yield (0.151 g dry cell weight/g glucose vs. 0.145 g dry cell weight/g glucose, or 0.184 Cmmol vs. 0.177 Cmmol). As was observed previously in the shake flask experiments described above, the overall difference in ethanol yield from glucose was not large. The SUT4 strain had a slightly higher (~1.11 fold) ethanol yield on glucose (0.372 g ethanol/g glucose or 0.485 Cmmol) than the control strain (0.335 g ethanol/g glucose or 0.437 Cmmol/Cmmol glucose).

The major difference between these two strains is in their kinetic rates of glucose and xylose consumption. The SUT4 strain consumes glucose at a much higher rate than the control strain (FIG. 5). In fact, the SUT4 strain consumed all of the glucose and xylose in the fermentation, while the URA3 control strain consumed approximately 88% of the glucose and did not consume enough xylose to significantly change the xylose concentration during the length of the fermentation.

The specific rate of glucose consumption for the SUT4 strain (0.284 g glucose gDCW⁻¹ h⁻¹ or 9.448 Cmmol gDCW⁻¹ h⁻¹) was 3.04 fold higher than that of the URA3 control strain (0.093 g Glucose gDCW⁻¹ h⁻¹ or 3.106 Cmmol gDCW⁻¹ h⁻¹). The specific rates of xylose consumption could not be compared due to the fact that the URA3 control strain had not significantly begun to utilize the xylose. Further bioreactor trials will be necessary to compare these rates.

The SUT4 and control strains also exhibited different rates of ethanol production. FIG. 6. shows a plot of ethanol production and glucose consumption. It should be noted that a small amount of ethanol produced by the SUT4 overexpression strain was due to the xylose consumed; however the rates and yields were calculated using the glucose only phase of fermentation. The specific rate of ethanol production for the SUT4 overexpression strain (0.106 g ethanol gDCW⁻¹ or 4.580 Cmmol gDCW⁻¹ h⁻¹) was 3.38 fold higher than that of the control strain (0.031 g ETOH gDCW⁻¹ h⁻¹ or 1.356 Cmmol gDCW⁻¹ h⁻¹).

Creation of the Δ pho13 Background Strains for Transporter Testing

In order to test the effect of putative transporters on xylose utilization, a robust *Saccharomyces cerevisiae* background that is capable of utilizing xylose in some capacity needed to be built. Previous research has shown that either the overexpression of TAL1 or the deletion of PHO13 promotes growth on xylose when the *Pichia stipitis* xylulokinase gene is expressed at a high level (10). The PHO13 deletion was chosen for use in these strains, as more detailed kinetic data during aerobic and low oxygen cultivations is available for a strain created in a similar genetic background (Van Vleet et al. (2008) Metab. Eng. Doi10.1016/j.ymben.2007.12.002).

The strains CEN.PK.111-27B and CEN.PK.102-3A were chosen as the genetic backgrounds in which to create these deletion strains (Entian K, Kötter P, (2007) 25 Yeast Genetic Strain and Plasmid Collections. In: Methods in Microbiology; Yeast Gene Analysis—Second Edition, Vol. Volume 36 (Ian Stansfield and Michael JR Stark ed), pp 629-666. Academic Press). The CEN.PK.111-27B carries the *trp1* and *leu2* mutations and the CEN.PK.102-3A strain carries the *ura3* and *leu2* mutations. The deletion of the PHO13 in these backgrounds will allow for the putative transporters to be tested in the presence of the xylose utilization genes expressed moderately and, separately, at a high level. These transporters will also be able to be tested at 2 different levels of expression. Moderate expression of the transporters will be possible using a centromeric LEU2 plasmid and high levels of expression will be possible using a multicopy LEU2 plasmid with a 2 μ m origin of replication.

The purified deletion cassette was then transformed into the CEN.PK strains using a standard lithium acetate method (Gietz & Woods (2002) Methods Enzymol 350, 87-96)). Out-growth was performed after the transformation for 3 hours in YPD liquid media, and then transformants were selected for on YPD plates containing 250 mg/ml G418. After 2 days of growth, growth of putative knockouts was confirmed by re-streaking on fresh G418 plates. These strains were then genotyped using primers that were designed 600 bp upstream and downstream of the PHO13 open reading frame. These primers amplify a product in both deletion and non deletion strains, however in deletion strains the product is approximately 650 bp larger than in the non deletion strain. Multiple transformants for each strain were identified as containing the deletion. Transformants containing the deletion originating from CEN.PK.111-27B were designated FPL.JHV.002 and those originating from CEN.PK.102-3A were designated FPL.JHV.003.

The xylose utilization genes (XYL1, XYL2, and XYL3) under the control of the strong constitutive TDH3 promoter were then introduced to the strains via plasmid. FPL.JHV.002 was transformed with the centromeric vector pRS314-X123 (10) and designated FPL.JHV.004. At the same time, FPL.JHV.003 was transformed with the multicopy vector pYES2-X123 (10) and designated FPL.JHV.005. Both of the vector bearing strains carry a *leu2* marker to allow for testing of the putative transporters. The plasmids and yeast strains are described in Table 6.

TABLE 5

Primers Used		
Primer Name	Function	Sequence
oPPL.JHV.021	PHO13F + HpaI	GATTACATGTTAACGATTGTTTCGACGCAACTACCC (SEQ ID NO: 8)
oPPL.JHV.022	PHO13R + HpaI	GATTACATGTTAACCTTCCCCAACAAGACCGAATTG (SEQ ID NO: 9)
oPPL.JHV.023	PHO13 Confirmation F	TTAAAACAAGAATTGGGGAG (SEQ ID NO: 10)
oPPL.JHV.024	PHO13 Confirmation	AAAGGTCTAATTATTCAATTTATCGAC (SEQ ID NO: 11)

The PHO13 deletion cassette was amplified out of the genomic DNA of *S. cerevisiae* CMB.JHV.pho13a (Van Vleet et al. (2008) Metab. Eng. Doi10.1016/j.ymben.2007.12.002) using the high fidelity PfuII polymerase (NEB, Finnzymes) and primers designed 400 bp upstream and downstream of the PHO13 open reading frame. This cassette contains the KANMX gene and has been shown previously to confer G418 resistance to PHO13 knockout strains in *S. cerevisiae* (Van Vleet et al. (2008) Metab. Eng. Doi10.1016/j.ymben.2007.12.002).

HpaI sites were added onto the ends of this amplified fragment for easy excision from a storage vector. The primers used in strain creation and confirmation are listed in Table 5. The amplified fragment was ligated into the pCR4 Blunt-TOPO vector and then transformed into TOP10 *E. coli* by a heat shock method. Transformants were selected for on kanamycin plates and digest checked to confirm the size of the insert. After verification, 100 μ g of plasmid DNA was prepared using a Qiagen maxiprep and digested with HpaI to release the deletion cassette. The deletion cassette was then gel purified.

Assessment of Putative Transporters by Whole Genome Expression Array Technology.

Cells of *Pichia stipitis* were cultivated in a bioreactor with either xylose or glucose as carbon sources along with mineral nutrients and urea for a nitrogen source using the medium of Verduyn et al. (16). Cells were grown either under fully aerobic or oxygen limited conditions. Cells were harvested, their mRNA was extracted and analyzed using a whole-genome expression array (NimbleGen). Transcripts corresponding to putative sugar transporters were identified and annotated. (Results not shown). Transcripts for seven genes (HXT2.4, HXT4, SUC1.5, XUT1, SUT4, XUT7 and YBR2) were induced to a higher level in cells grown on xylose than in cells grown on glucose. All but SUT4 were induced to their highest levels on xylose under oxygen limiting conditions. Transcripts for seven genes (AUT1, HGT2, HUT1, MAL4, RGT2, STL1 and SUT1) were all induced to their highest levels when cells were cultivated on glucose. Transcript for one gene (SUT1) was induced to its highest level under oxygen limitation; the other six were induced to their highest

levels under aerobic conditions. The specific induction of transcripts on xylose under aerobic or oxygen limited conditions was taken as an indicator of the effective function of this gene when cells were cultivated on xylose. Of these genes, SUT1, SUT2 and SUT3 were previously recognized and described as glucose/xylose facilitator transport proteins.

It is to be understood that the invention is not limited in its application to the details of construction and the arrangement of components set forth in the following description. The invention is capable of other embodiments and of being practiced or of being carried out in various ways. Also, it is to be understood that the phraseology and terminology used herein is for the purpose of description and should not be regarded as limiting. The use of "including," "comprising," or "having"

and variations thereof herein is meant to encompass the items listed thereafter and equivalents thereof as well as additional items.

It also is understood that any numerical range recited herein includes all values from the lower value to the upper value. For example, if a concentration range is stated as 1% to 50%, it is intended that values such as 2% to 40%, 10% to 30%, or 1% to 3%, etc., are expressly enumerated in this specification. These are only examples of what is specifically intended, and all possible combinations of numerical values between and including the lowest value and the highest value enumerated are to be considered to be expressly stated in this application.

 SEQUENCE LISTING

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<211> LENGTH: 1701

<212> TYPE: DNA

<213> ORGANISM: *Pichia stipitis*

<220> FEATURE:

<223> OTHER INFORMATION: xylose transporter XUT1

<400> SEQUENCE: 1

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gacagagaag acgaatgtht gtccgttctt tccaacttgc gttccttgag taaggaagat      780
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ttcaaccaat acaagtccat gattactcac tacccaacct tcaagcgtgt tgcagttgcc      960
tgtttaatta tgacctcca acaatggact ggtgttaact tcatcttgta ctatgctcca     1020
ttcatcttca gttctttagg tttgtctgga aacaccattt ctcttttagc ttctgggtgt     1080
gtcggtagtc tcattgttct gtctaccatt ccagctgttc tttgggtctg cagacttggt     1140
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gcaatcttag gtcagttcgg tggttaactt gtcaaccact ccggtgctgg ttgggttgct     1260
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<210> SEQ ID NO 2
<211> LENGTH: 566
<212> TYPE: PRT
<213> ORGANISM: Pichia stipitis
<220> FEATURE:
<223> OTHER INFORMATION: xylose transporter XUT1

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<400> SEQUENCE: 2

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20          25          30
Asn Ser Arg Ser Phe Phe Ile Ala Val Phe Ala Ser Leu Gly Gly Leu
35          40          45
Val Tyr Gly Tyr Asn Gln Gly Met Phe Gly Gln Ile Ser Gly Met Tyr
50          55          60
Ser Phe Ser Lys Ala Ile Gly Val Glu Lys Ile Gln Asp Asn Pro Thr
65          70          75          80
Leu Gln Gly Leu Leu Thr Ser Ile Leu Glu Leu Gly Ala Trp Val Gly
85          90          95
Val Leu Met Asn Gly Tyr Ile Ala Asp Arg Leu Gly Arg Lys Lys Ser
100         105         110
Val Val Val Gly Val Phe Phe Phe Phe Ile Gly Val Ile Val Gln Ala
115         120         125
Val Ala Arg Gly Gly Asn Tyr Asp Tyr Ile Leu Gly Gly Arg Phe Val
130         135         140
Val Gly Ile Gly Val Gly Ile Leu Ser Met Val Val Pro Leu Tyr Asn
145         150         155         160
Ala Glu Val Ser Pro Pro Glu Ile Arg Gly Ser Leu Val Ala Leu Gln
165         170         175
Gln Leu Ala Ile Thr Phe Gly Ile Met Ile Ser Tyr Trp Ile Thr Tyr
180         185         190
Gly Thr Asn Tyr Ile Gly Gly Thr Gly Ser Gly Gln Ser Lys Ala Ser
195         200         205
Trp Leu Val Pro Ile Cys Ile Gln Leu Val Pro Ala Leu Leu Leu Gly
210         215         220
Val Gly Ile Phe Phe Met Pro Glu Ser Pro Arg Trp Leu Met Asn Glu
225         230         235         240
Asp Arg Glu Asp Glu Cys Leu Ser Val Leu Ser Asn Leu Arg Ser Leu
245         250         255
Ser Lys Glu Asp Thr Leu Val Gln Met Glu Phe Leu Glu Met Lys Ala
260         265         270
Gln Lys Leu Phe Glu Arg Glu Leu Ser Ala Lys Tyr Phe Pro His Leu
275         280         285
Gln Asp Gly Ser Ala Lys Ser Asn Phe Leu Ile Gly Phe Asn Gln Tyr
290         295         300
Lys Ser Met Ile Thr His Tyr Pro Thr Phe Lys Arg Val Ala Val Ala
305         310         315         320

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<210> SEQ ID NO 4

<211> LENGTH: 550

<212> TYPE: PRT

<213> ORGANISM: Pichia stipitis

<220> FEATURE:

<223> OTHER INFORMATION: xylose transporter SUT4, PsSUT4, glucose/xylose facilitator PsSut4

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20        25        30
Gln Ser Asp Leu Ala Ser Ile Pro Ala Thr Gly Ile Lys Ala Tyr Leu
35        40        45
Leu Val Cys Phe Phe Cys Met Leu Val Ala Phe Gly Gly Phe Val Phe
50        55        60
Gly Phe Asp Thr Gly Thr Ile Ser Gly Phe Leu Asn Met Ser Asp Phe
65        70        75        80
Leu Ser Arg Phe Gly Gln Asp Gly Ser Glu Gly Lys Tyr Leu Ser Asp
85        90        95
Ile Arg Val Gly Leu Ile Val Ser Ile Phe Asn Ile Gly Cys Ala Ile
100       105       110
Gly Gly Ile Phe Leu Ser Lys Ile Gly Asp Val Tyr Gly Arg Arg Ile
115       120       125
Gly Ile Ile Ser Ala Met Val Val Tyr Val Val Gly Ile Ile Ile Gln
130       135       140
Ile Ser Ser Gln Asp Lys Trp Tyr Gln Leu Thr Ile Gly Arg Gly Val
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caaaagtgtt cccttaaacc agtttagact gaaacaaccg aatgtaatca gggtagatga    240
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<212> TYPE: DNA

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<400> SEQUENCE: 6

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gtcatccttt ccactcccat tacaaaacct cattacatct cattagtcaa agtccactca    780
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<212> TYPE: DNA
<213> ORGANISM: Pichia stipitis

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 <220> FEATURE:
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<400> SEQUENCE: 11

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 <212> TYPE: DNA
 <213> ORGANISM: Pichia stipitis
 <220> FEATURE:
 <223> OTHER INFORMATION: xylose transporter PsSUT2, SUT2

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We claim:

1. A method of producing ethanol or xylitol from the fermentation of xylose comprising: culturing a yeast strain in contact with a xylose-containing material under suitable conditions for a period of time sufficient to allow fermentation of at least a portion of the xylose to ethanol or xylitol, wherein the yeast strain comprises a nucleic acid comprising a coding sequence operably linked to a promoter not natively associated with the coding sequence, the coding sequence encoding a glucose/xylose transporter polypeptide having at least 95% amino acid identity to SEQ ID NO:2, or at least 95% amino acid identity to SEQ ID NO:4 and comprising a threonine residue at the amino acid position corresponding to position 544 of SEQ ID NO:4.

2. The method of claim 1, wherein the strain exhibits increased uptake of xylose relative to a control yeast lacking the nucleic acid.

3. The method of claim 1, wherein at least a portion of the xylose is converted to ethanol.

4. The method of claim 1, wherein the xylose-containing material further comprises glucose.

5. The method of claim 1, wherein the promoter is a constitutive promoter or promoter inducible under oxygen limiting conditions.

6. The method of claim 1, wherein the polypeptide comprises SEQ ID NO:2.

7. The method of claim 1, wherein the coding sequence encodes a glucose/xylose transporter with 95% amino acid identity to SEQ ID NO:2 and a glucose/xylose transporter polypeptide having at least 95% amino acid identity to SEQ ID NO:4 and comprising a threonine or serine residue at the amino acid position corresponding to position 544 of SEQ ID NO:4.

8. The method of claim 1, wherein the polypeptide comprises SEQ ID NO:4.

9. The method of claim 1, wherein the yeast strain is a *Pichia stipitis* or *Saccharomyces*.

10. The method of claim 1, wherein at least a portion of the xylose is converted to xylitol.

11. The method of claim 10, wherein the strain exhibits increased xylitol production relative to a control yeast lacking the nucleic acid.

12. The method of claim 10, wherein the strain has reduced xylitol dehydrogenase activity such that xylitol accumulates.

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